

ANALYST:		VPDES NO.	
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Parameter: Biochemical Oxygen Demand  
Method: Dissolved Oxygen Depletion  
1/08

METHOD OF ANALYSIS:

	18th Edition of Standard Methods 5210 B
	21 <sup>st</sup> or On-Line Edition of Standard Methods 5210 B (01)

- BOD is a method-defined analyte so modifications are not allowed. [40 CFR Part 136.6]**
- 1) Is a certificate of operator competence or initial demonstration of capability available for each analyst/operator performing this analysis? **NOTE:** Analyze 4 samples of known BOD concentration with each sample having +/- 20 % recovery. [SM 1020 B.1]
  - 2) Are incubation bottles cleaned well with detergent, rinsed thoroughly, and drained before use? **NOTE:** Do not use anything to clean bottles that will inhibit growth of seed material such as bleach or chromic acid. [2.a]
  - 3) Are nutrient solutions (calcium chloride, magnesium sulfate, ferric chloride, and phosphate buffer) free of biological growths and solids, and within shelf lives? [3]
  - 4) Is the phosphate buffer solution documented to be at pH 7.2 when prepared? [3.a]
  - 5) Are all nutrient solutions added at a rate of 1 mL/L to dilution water? **NOTE:** Hach slurry pillows are acceptable. [18<sup>th</sup> ed. 4.a; 21<sup>st</sup> ed. 5.a]
  - 6) Is dilution water free of contamination or growths? [3.a]
  - 7) Are chlorinated samples checked for chlorine using an appropriate method? **NOTE:** The chlorine check must be documented. [18<sup>th</sup> ed. 4.e.2; 21<sup>st</sup> ed. 4.b.2]
  - 8) Are samples containing residual chlorine, dechlorinated with sodium sulfite? [18<sup>th</sup> ed. 4.e.2; 21<sup>st</sup> ed. 4.b.2]
  - 9) Is sodium sulfite dechlorinating solution prepared fresh daily? Documentation needed. [3.f]
  - 10) Are samples checked for caustic alkalinity or acidity? (pH <6.0 or >8.0 SU) [18<sup>th</sup> ed. 4.e.1; 21<sup>st</sup> ed. 4.b.1]
  - 11) Are samples containing acidity or caustic alkalinity adjusted to fall between pH 6.5 and 7.5 for 18<sup>th</sup> ed. and 7.0 and 7.2 for 21<sup>st</sup> ed.? [18<sup>th</sup> ed. 4.e.2; 21<sup>st</sup> ed. 4.b.2]
  - 12) If the initial DO exceeds saturation at 20°C, is sample stripped of excess DO by agitation or aeration? [18<sup>th</sup> ed. 4.e.4; 21<sup>st</sup> ed. 4.b.4]
  - 13) Are sample initial dissolved oxygen concentrations between 7 mg/L and saturation? [18<sup>th</sup> ed. 6.b; 21<sup>st</sup> ed. 8.b]
  - 14) Are samples allowed to reach 20 ± 1°C (18<sup>th</sup> ed.) or 20 ± 3°C (21<sup>st</sup> ed.) before making dilutions? **NOTE:** Documentation is necessary. [18<sup>th</sup> ed. 4.e.5; 21<sup>st</sup> ed. 5.b]
  - 15) Is dilution water saturated with dissolved oxygen at 20°C before use? [18<sup>th</sup> ed. 4.a; 21<sup>st</sup> ed. 5.a]
  - 16) If seeding is necessary for samples being analyzed is appropriate seed material used? [18<sup>th</sup> ed. 4.d.1; 21<sup>st</sup> ed. 5.d]
  - 17) Is a seed control series run for seeded samples? [18<sup>th</sup> ed. 4.d.2; 21<sup>st</sup> ed. 6.d]

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18) For seeded samples, is the calculated seed correction between 0.6 and 1.0 mg/L? [18 <sup>th</sup> ed. 4.d.2; 21 <sup>st</sup> ed. 5.d]		
19) If CBOD <sub>5</sub> is analyzed, are dilutions and seed correction series inhibited with 2.2% 2-chloro 6-trichloro-methyl pyridine (HACH nitrification inhibitor 2533 or equivalent)? <b>NOTE:</b> Polyseed-NX is not approved for use in VPDES testing. Do not add inhibitor to GGA. (For exception see question 36.) A separate seed correction series without inhibitor must also be analyzed so that GGA can be properly calculated. [18 <sup>th</sup> ed. 4.e.6; 21 <sup>st</sup> ed. 5.e.1]		
20) Are samples prepared without entraining air in BOD bottles? [18 <sup>th</sup> ed. 4.f.2; 21 <sup>st</sup> ed. 5.c.1]		
21) Are dissolved oxygen concentrations measured correctly (see Winkler/Azide, LDO, or DO electrode Checklists)? [18 <sup>th</sup> ed. 4.e.6; 21 <sup>st</sup> ed. 5.g]		
22) Are water seals maintained? [18 <sup>th</sup> ed. 4.f.2; 21 <sup>st</sup> ed. 5.f]		
23) Are samples incubated in the dark at 20 ± 1°C for 5 days? <b>NOTE:</b> Documentation is necessary. [18 <sup>th</sup> ed. 4.f.2; 21 <sup>st</sup> ed. 5.h]		
24) Is the final DO of at least one dilution at least 1 mg/L after 5 days? [18 <sup>th</sup> ed. 5; 21 <sup>st</sup> ed. 6.a]		
25) Is the DO depletion of at least one dilution at least 2 mg/L after 5 days? (Disregard if sample is not diluted) [18 <sup>th</sup> ed. 4.f; 21 <sup>st</sup> ed. 6.a]		
26) Are all bottles meeting the depletion criteria averaged for final BOD results? [18 <sup>th</sup> ed. 5; 21 <sup>st</sup> ed. 7]		
27) Is a dilution water blank run for each test series and are blank depletions recorded on bench sheets? [18 <sup>th</sup> ed. 4.h; 21 <sup>st</sup> ed. 6.c]		
28) Is the dilution water blank DO depletion consistently less than 0.4 mg/L (DEQ criterion)? [18 <sup>th</sup> ed. 4.h; 21 <sup>st</sup> ed. 6.c]		
29) Are dilutions capable of demonstrating permit excursions? [18 <sup>th</sup> ed. 4.f; 21 <sup>st</sup> ed. 5.c]		
30) Are at least three dilutions analyzed for each sample? [18 <sup>th</sup> ed. 4.f; 21 <sup>st</sup> ed. 5.c]		
31) Are sample results calculated correctly? [18 <sup>th</sup> ed. 5; 21 <sup>st</sup> ed. 7.c]		
<p>BOD (mg/L) = <math>\frac{(D1 - D2) - (B1 - B2)f}{P}</math> where</p> <p>D1 = D.O. of diluted sample after preparation  D2 = D.O. of diluted sample after 5 days  P = decimal volumetric fraction of sample  B1 = D.O. of seed control before incubation  B2 = D.O. of seed control after incubation  f = ratio of seed in sample to seed in control (% seed in D1)/(% seed in B1)</p>		
32) Are BOD bottles (blank/seed/sample/GGA) chosen at random? [Permit]		
33) Is the glucose-glutamic acid (GGA) check run at least once each week of analysis? [18 <sup>th</sup> ed. 4.c; 21 <sup>st</sup> ed. 6.b]		
34) Is the GGA prepared immediately before use? [3.h]		
35) Is the BOD <sub>5</sub> of the 2% dilution of the GGA standard within the range of 198 ± 30.5 mg/L? <b>NOTE:</b> 21 <sup>st</sup> ed. requires set up of three test bottles and the average result be within range. [18 <sup>th</sup> ed. 6; 21 <sup>st</sup> ed. 6.b]		
36) If citing the 21 <sup>st</sup> ed. is nitrification inhibitor added to GGA test bottles if seed is obtained from a source that is nitrifying? [21 <sup>st</sup> ed. 6.b]		
37) Is data flagged on benchsheet and DMR when QC problems occur (e.g., GGA out of range, blank >0.4 mg/L, bubbles in bottle at end of incubations)? [Permit & 21 <sup>st</sup> ed. 7.b]		

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38) Is raw data evaluated to determine if toxicity is present? [18 <sup>th</sup> ed. 5; 21 <sup>st</sup> ed. 7.b]		
39) If toxicity is present, are BOD results reported properly? [18 <sup>th</sup> ed. 6; 21 <sup>st</sup> ed. 6.b]		
40) Is a duplicate sample analyzed after every 20 samples if citing 18 <sup>th</sup> ed. [1020 B.6] or weekly for 21 <sup>st</sup> ed. [2540 D.3.c]? <b>NOTE:</b> "Duplicate sample" must have same dilutions as "sample".		
41) If duplicate sample is analyzed, is the relative percent difference (RPD) $\leq 20$ ? [18 <sup>th</sup> ed. Table 1020 I; 21 <sup>st</sup> ed. DEQ]		
42) Is a laboratory control sample (LCS) analyzed at the required frequency? [18 <sup>th</sup> ed. 1020 B. 3; 21 <sup>st</sup> ed. 11020 C.1]		

PROBLEMS: